

## Mechanism of Ultra-Fast Actin-Myosin Sliding Producing Cytoplasmic Streaming in Giant Algal Cell, Studied Using the Centrifuge Microscope

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### Abstract

In giant intermodal cells of green algae *Chara collaria*, cytoplasmic streaming is produced by ATP-dependent sliding between myosin heads extending from amorphous cytoplasmic organelles and actin filament arrays (actin cables) fixed on chloroplast rows. The velocity of cytoplasmic streaming is many times faster than the maximum myofibril sliding in skeletal muscle. In this article, we compared steady-state force-velocity (P-V) relations between cytoplasmic myosin and skeletal and cardiac muscle myosins using the centrifuge microscope, in which myosin-coated latex beads were made to slide along the actin cables under various centrifugal forces. In contrast with the hyperbolic P-V relation of actin-myosin sliding in skeletal and cardiac myosins, the P-V relation of cytoplasmic myosin versus actin cable sliding was a straight line, indicating a very large duty ratio and a very small rate of chemo-mechanical energy conversion. Possible mechanisms of the ultra-fast actin-myosin sliding are discussed.

### Highlights

- The velocity of cytoplasmic streaming, caused by ATP-dependent sliding between cytoplasmic myosin and actin cables in giant algal cells is many times faster than ATP-dependent actin-myosin sliding in skeletal and cardiac muscles.
- The mechanism of ultra-fast actin-myosin sliding was studied using the centrifuge microscope, in which beads coated with cytoplasmic myosin were made to slide along actin cables under various centrifugal forces serving as loads against cytoplasmic myosin versus actin cable sliding.
- Unlike the hyperbolic force-velocity (P-V) relation of skeletal and cardiac muscle actin-myosin sliding, the P-V relation of cytoplasmic actin myosin sliding was a straight line irrespective of the force generated by cytoplasmic myosin.
- These results indicate a very large duty ratio and a very small efficiency of chemo-mechanical energy conversion in cytoplasmic actin-myosin sliding.

**Keywords:** Ultra-fast actin-myosin sliding; Cytoplasmic streaming; Giant algal cell; Force-velocity relation; Centrifuge microscope; Maximum shortening velocity

### Characteristic Features of ATP-Dependent Actin-Myosin Interaction Producing Cytoplasmic Streaming

Cytoplasmic streaming is widely observed in plant cells. The mechanism of cytoplasmic streaming has been investigated on giant intermodal cells of green algae *Chara collaria* (diameter, ~0.5 mm; length, >10 cm) (Figure 1) with the following results: (1) The motive force for *Chara* cytoplasmic streaming is generated by ATP-dependent interaction between *Chara* cytoplasmic myosin molecules extending from amorphous cytoplasmic organelles and well-organized straight actin filament arrays fixed on chloroplast rows, called actin cables (Figure 2); (2) The direction of streaming is reversed across the indifferent zone where chloroplast is absent, reflecting the reversal of actin filament polarity across the indifferent zone; (3) Since the two indifferent zones run parallel to the intermodal cell long axis, cytoplasmic streaming rotates longitudinally within the cell [1-7].

Although both cytoplasmic streaming and muscle contraction are produced by ATP-dependent actin-myosin sliding, and regulated by intracellular Ca ion concentration, skeletal muscle is relaxed at  $pCa \geq 7$  and fully contracts at  $pCa \leq 5$ , while cytoplasmic streaming occurs at  $pCa \geq 7$ , and stops at  $pCa \leq 6$  [8-10]. If small beads coated with skeletal muscle myosin is made to slide along actin cables in the presence of ATP, the bead movement is insensitive to  $pCa$  [11], indicating the absence of regulatory proteins in actin cables. On the other hand,

the stoppage of cytoplasmic streaming is caused by tight cytoplasmic myosin-actin cable linkages, which is stronger than their rigor linkages in the absence of ATP, suggesting that *Chara* cytoplasmic myosin can bind with actin cables in two different modes.

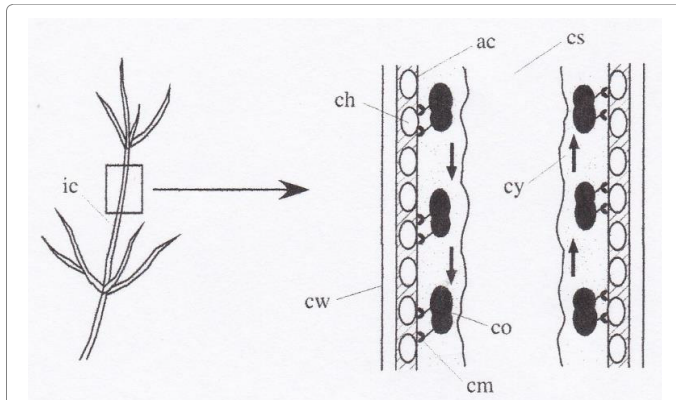
The most interesting feature of cytoplasmic streaming in giant intermodal cells is that its velocity ( $\geq 50 \mu\text{m/s}$ ) [1] is many times larger than the maximum velocity of skeletal muscle shortening, i.e. sliding between actin and myosin filaments ( $\leq 10 \mu\text{m/s}$ ). In the presence of ATP, beads coated with skeletal muscle myosin move on actin cables at velocities similar to that of actin-myosin sliding in muscle [12,13]. In contrast, skeletal muscle actin filaments move on a glass surface coated with *Chara* cytoplasmic myosin at velocities similar to that of native cytoplasmic streaming [14]. These results indicate that it is *Chara* cytoplasmic myosin that causes the ultra-fast velocity of cytoplasmic

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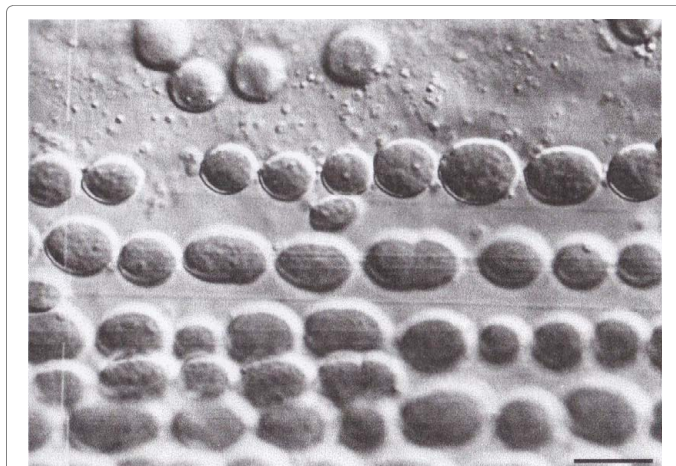
**Received** January 10, 2017; **Accepted** February 13, 2018; **Published** February 23, 2018

**Citation:** Chaen S, Sugi H (2018) Mechanism of Ultra-Fast Actin-Myosin Sliding Producing Cytoplasmic Streaming in Giant Algal Cell, Studied Using the Centrifuge Microscope. J Material Sci Eng 7: 426. doi: [10.4172/2169-0022.1000426](https://doi.org/10.4172/2169-0022.1000426)

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**Figure 1:** Diagram showing cytoplasmic streaming in giant intermodal cell of green algae *Chara corallina*. ac, actin cables; ch, chloroplast; cm, cytoplasmic myosin; co, cytoplasmic organelle; cs, cell sap; cw, cell wall; cy, streaming cytoplasm, Arrows indicate direction of streaming; ic, intermodal cell.



**Figure 2:** Actin cables on the inner surface of chloroplast rows in an intermodal cell of *Chara corallina*. Calibration bar, 10  $\mu\text{m}$  [7].

streaming. As with skeletal muscle myosin, *Chara* cytoplasmic myosin molecule is a dimer consisting of two heavy chains with one head and one tail, so that they have two heads and one helical tail, but the tail length is much shorter than that in skeletal muscle myosin molecules [15].

### Centrifuge Microscope to Study Steady-state Force-velocity Relation of ATP-dependent Sliding between Myosin and Actin Cables

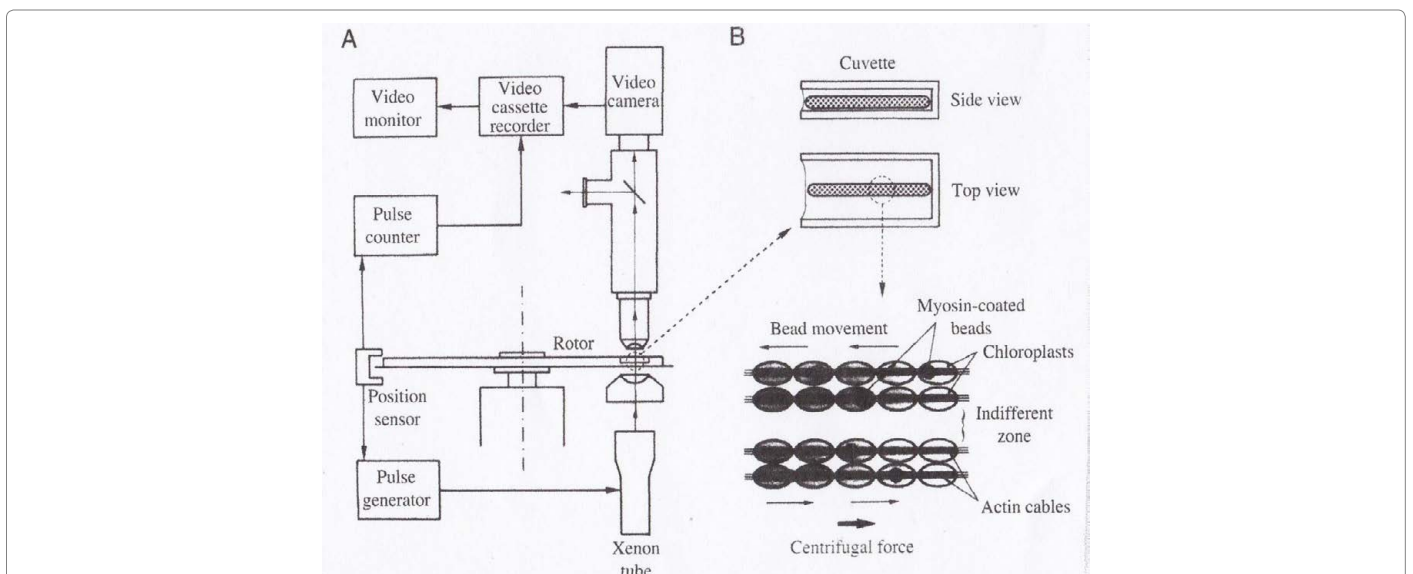
Figure 3 illustrates the setup of the centrifuge microscope constructed in our laboratory [13]. It consisted of a light microscope, a rotor, and a stroboscopic light source. The specimen used was a short segment of an intermodal cell of green algae, into which myosin-coated polystyrene beads (diameter, 2.8  $\mu\text{m}$ ; specific gravity, 1.3) were introduced together with experimental solution containing MgATP. The specimen was placed in a cuvette, fixed on the rotor of the centrifuge microscope, in such a way that actin cables were parallel to the direction of centrifugal forces applied. Centrifugal forces directed opposite to the bead movement served as positive external loads on myosin molecules producing actin-myosin sliding. The amount of centrifugal force (=load)  $F$  on the bead is given by:

$$F = \Delta\rho V r \omega^2,$$

Where  $\Delta\rho$  is the difference in density between the bead and the surrounding medium (0.3  $\text{g cm}^{-3}$ ,  $V$  is the bead volume (12  $\mu\text{m}^3$ ),  $r$  is the effective radius of centrifugation (4.5-7 cm) and  $\omega$  is the angular velocity of the rotor. Further details of the centrifuge microscope have been described elsewhere [13]. Experiments were made at 20-23°C.

### Steady-state Force-velocity Relation of ATP-dependent Sliding between Skeletal Muscle Myosin and Actin Cables

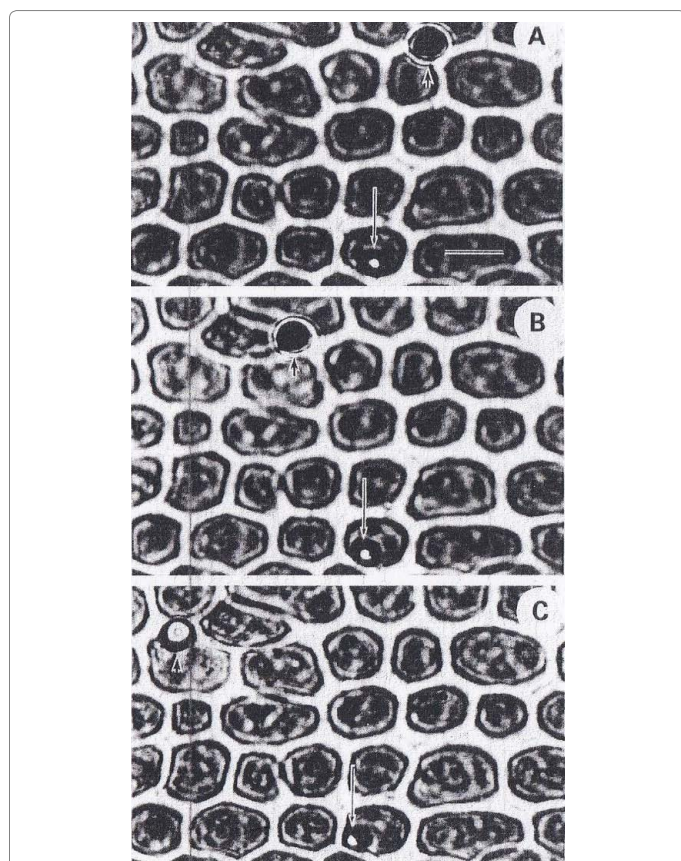
First, we will describe steady-state force-velocity relation of ATP-dependent sliding between skeletal muscle myosin, prepared from rabbit psoas muscle, and actin cables. Under a constant centrifugal force serving as a positive external load, myosin-coated beads are



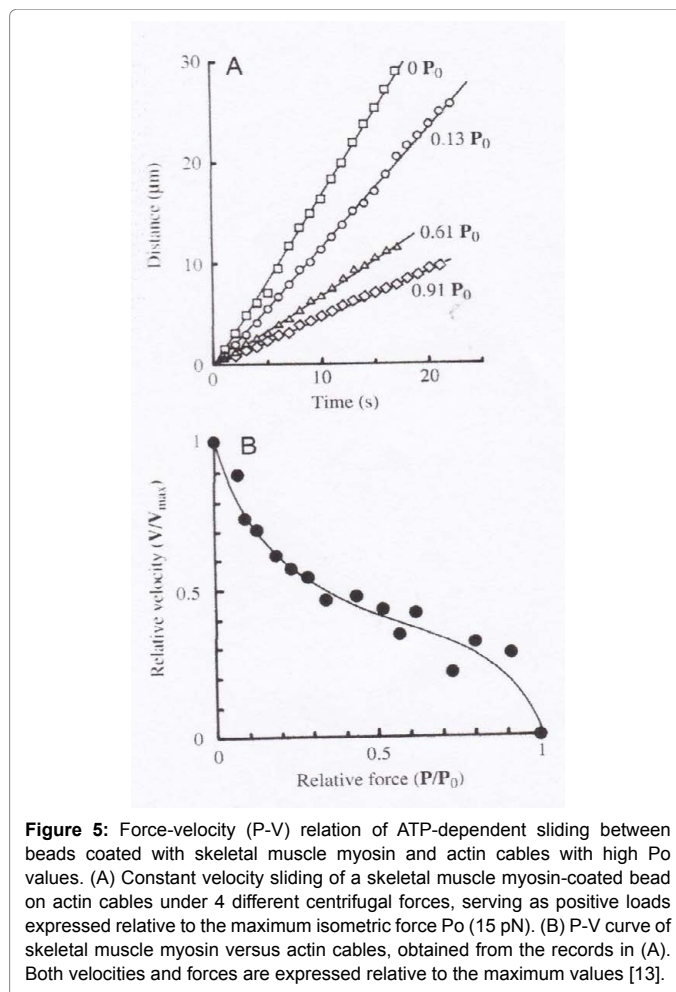
**Figure 3:** Centrifuge microscope for studying ATP-dependent sliding between myosin-coated beads and actin cables. (A) Diagram showing centrifuge microscope and video recording system. (B) Application of centrifugal forces serving as positive loads on the sliding beads [13].

observed to move along chloroplast rows (Figure 4) [13,16]. The beads continue moving with constant velocities determined by the amount of external load over a long distance, indicating the steady state between the load (=force  $P$ ), generated by myosin molecules on the bead and the velocity of actin-myosin sliding (Figure 5A). The maximum unloaded velocity of bead movement was 1.6-3.6  $\mu\text{m/s}$ . The bead stopped moving when the load was increased to the maximum isometric force  $P_0$  generated by myosin molecules on the bead. As shown in Figure 5B, the force-velocity ( $P$ - $V$ ) curve of the bead movement with high  $P_0$  values (10-39 pN) was hyperbolic in shape in the low force region, but deviated from hyperbola in the high force region. The shape of the  $P$ - $V$  curve was nearly similar to that obtained from tetanized single skeletal muscle fibers [17,18], and  $\text{Ca}^{2+}$ -activated single skinned muscle fibers (Figure 6) [19,20]. These results not only indicate that skeletal muscle myosin molecules on the bead retain their basic kinetic properties of contracting muscle, but also close similarities between actin cables and skeletal muscle actin filaments.

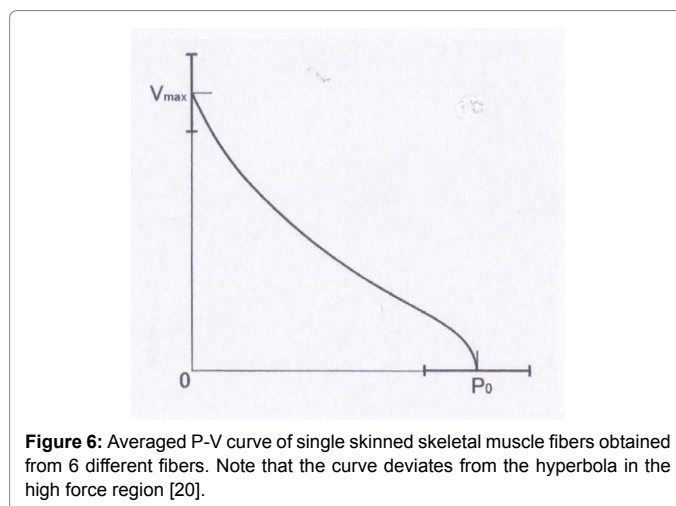
Meanwhile, in the  $P$ - $V$  curves of the bead movement with low  $P_0$  values (3.5-5.9 pN), the hyperbolic part at low force region became much less pronounced (Figure 7). Similar results were obtained on the  $P$ - $V$  curves of sliding movement of the beads coated with two different myosin isoforms, V1 and V3, obtained from rabbit cardiac muscle [21]. In this study, we made a tentative explanation for the increasing



**Figure 4:** Selected video frames showing two myosin-coated polystyrene beads sliding from right to left along chloroplast rows, on which actin cables (not visible) run straight. One bead (diameter, 4.5  $\mu\text{m}$ , upper edge of each frame, indicated by upward arrows) is coated with skeletal muscle myosin, while the other bead (diameter, 2.8  $\mu\text{m}$ , lower edge of each frame, indicated by downward arrows) are coated with skeletal muscle myosin previously treated anti-subfragment 2 antibody [16].



**Figure 5:** Force-velocity ( $P$ - $V$ ) relation of ATP-dependent sliding between beads coated with skeletal muscle myosin and actin cables with high  $P_0$  values. (A) Constant velocity sliding of a skeletal muscle myosin-coated bead on actin cables under 4 different centrifugal forces, serving as positive loads expressed relative to the maximum isometric force  $P_0$  (15 pN). (B)  $P$ - $V$  curve of skeletal muscle myosin versus actin cables, obtained from the records in (A). Both velocities and forces are expressed relative to the maximum values [13].



**Figure 6:** Averaged  $P$ - $V$  curve of single skinned skeletal muscle fibers obtained from 6 different fibers. Note that the curve deviates from the hyperbola in the high force region [20].

deviation of  $P$ - $V$  curves from the hyperbola with decreasing  $P_0$  in the following way. The ATP-dependent-actin-myosin sliding is caused by cyclic attachment and detachment between myosin heads and actin filaments. According to the Huxley contraction model [22], the probability that a myosin head is attached to actin is  $0 < p < 1$ , and the probability that all myosin heads interacting with actin are in detached state, i.e. off probability  $P_{off}$  is given by:

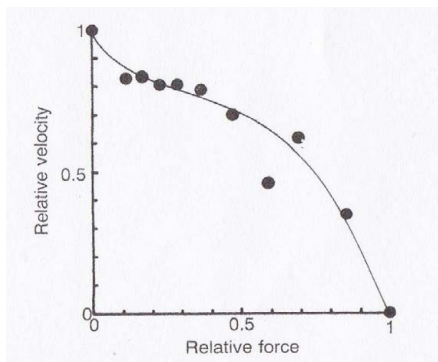
$$P_{off}=(1-p)^N,$$

where  $N$  is the number of myosin heads interacting with actin. As shown in Figure 8,  $P_{off}$  becomes substantial for small  $N$ . During the period of  $P_{off}$ , beads are displaced in the direction opposite to that of actin-myosin sliding, to result in retardation of the observed sliding velocity under the centrifuge microscope. Assuming the appropriate dependence of  $p$  on the amount of load for cardiac myosin isoforms V1 and V3, the shortening velocities in the small load (=force) region of P-V curves with small  $P_o$  values are corrected to be larger than the observed values. This implies that, if the number of myosin heads interacting with actin is small, the shortening velocity in the small load region is slowed down due to increase in the moment, at which all myosin heads detach from actin.

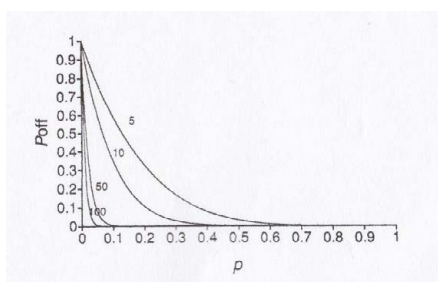
In Figure 9, broken lines indicate P-V curves for V1 myosin(A) and for V2 myosin actually obtained by the centrifuge microscope, while solid curves indicate P-V curves corrected on the assumption that  $P_{off}=0$  for all load (=force) region. It can be seen that the marked deviation from the hyperbola of the P-V curve with low  $P_o$  values (broken lines, cf. Figure 7) can be made much less pronounced (solid lines) to resemble the P-V curve with high  $P_o$  values (cf. Figure 5B).

### Steady-state Force-velocity Relation of ATP-dependent Sliding between Cytoplasmic Myosin and Actin Cables

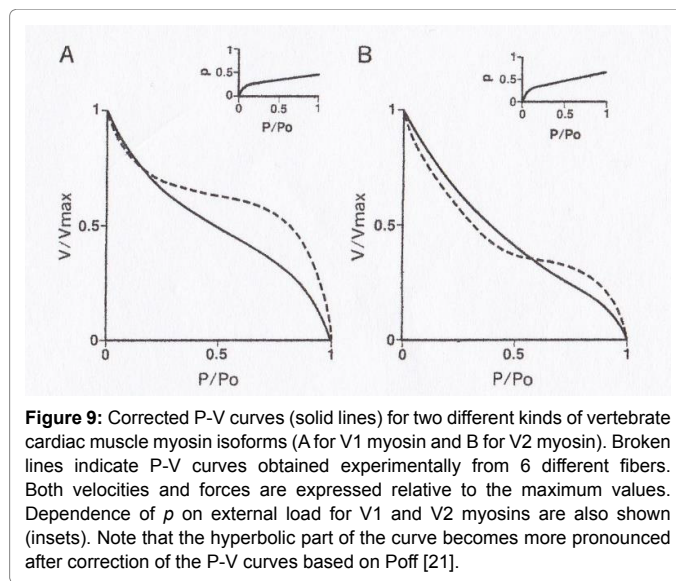
During the course of experiments with skeletal muscle myosin, we found that, when uncoated beads were introduced into the giant



**Figure 7:** P-V curve of skeletal muscle myosin versus actin cables with low  $P_o$  values. Both velocities and forces are expressed relative to the maximum values. Note that the hyperbolic part of the curve at low force region is much less pronounced compared to the P-V curve shown in Figure 5B [13].



**Figure 8:** Dependence of  $P_{off}$ , i.e. the probability that all myosin heads interacting with actin are detached at any chosen moment, on the fraction of time, in which a myosin head is attached firmly to actin within one cycle of attachment-detachment cycle  $p$  ( $0 < p < 1$ ). Numbers alongside each curve indicate number of myosin heads interacting with actin [21].



**Figure 9:** Corrected P-V curves (solid lines) for two different kinds of vertebrate cardiac muscle myosin isoforms (A for V1 myosin and B for V2 myosin). Broken lines indicate P-V curves obtained experimentally from 6 different fibers. Both velocities and forces are expressed relative to the maximum values. Dependence of  $p$  on external load for V1 and V2 myosins are also shown (insets). Note that the hyperbolic part of the curve becomes more pronounced after correction of the P-V curves based on  $P_{off}$  [21].

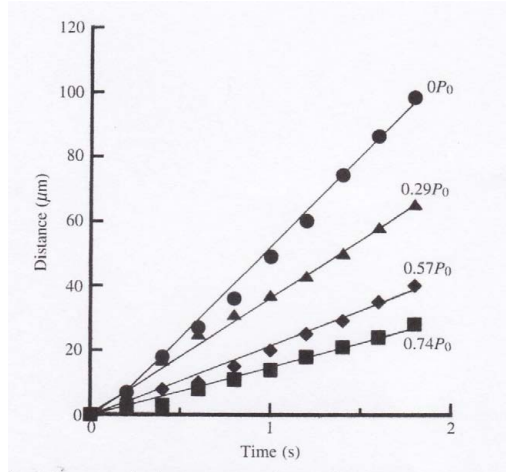
internodal cell preparation, the bead always started moving along actin cables with velocities similar to those of native cytoplasmic streaming. This phenomenon was explained as being due to *Chara* cytoplasmic myosin molecules, which had been detached from cytoplasmic organelles and remained in the internodal cell preparation; such *Chara* myosin molecules attach to the uncoated bead surface, in such a way to produce cytoplasmic streaming in the same manner when they attach to cytoplasmic organelles. Therefore, we took the opportunity to study P-V curves of sliding movement of *Chara* myosin-coated beads along actin cables [23].

As has been the case in skeletal and cardiac muscle myosins, beads spontaneously covered with *Chara* cytoplasmic myosin, which will hereafter be simply called cytoplasmic myosin, moved with constant velocities under various centrifugal forces serving as external loads, indicating a definite steady-state between the force (=load) generated by cytoplasmic myosin and the velocity of actin-myosin sliding (Figure 10). The velocity of bead movement decreased with increasing external load, and the bead eventually stopped moving when the load became equal to the maximum isometric force  $P_o$  generated by cytoplasmic myosin molecules. The beads that stopped moving remained in the same position on actin cables for 5-10 s and then suddenly detached from actin cables to flow away out of microscopic field. The value of  $P_o$  showed a wide range of variation from 1.0 to 13 pN (mean  $\pm$  SD, 6.2  $\pm$  4.5 pN,  $n=12$ ).

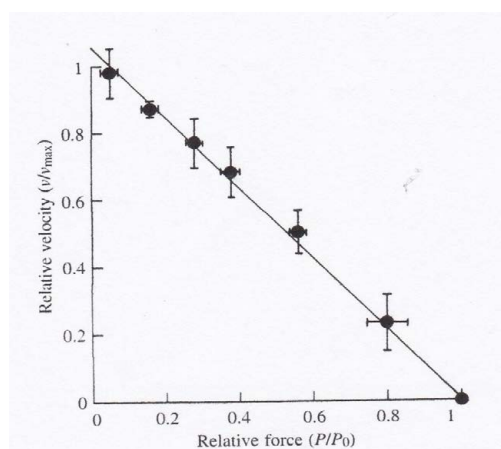
The steady-state P-V curves of the ATP-dependent bead movement along actin cables were obtained either by gradually increasing external load or by randomly changing external load with similar results. A typical example of the P-V relation obtained from a bead ( $P_o=13$  pN) is shown in Figure 11 (filled circles). The P-V curve was nearly straight in shape. The viscous drag force  $F$  on the bead can be calculated as:

$$F=6\pi\eta r v,$$

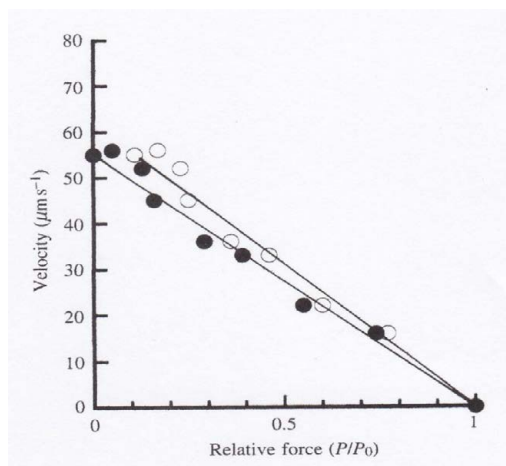
Where  $\eta$  is the viscosity of the medium ( $0.001$  Ns/m<sup>2</sup>),  $r$  is the bead radius ( $1.4$   $\mu$ m) and  $v$  is the bead velocity (in  $\mu$ m/s). For  $v=60$   $\mu$ m/s, for example, the drag force is 1.6 pN, amounting to more than 10% of measured value of  $P_o$ . In Figure 11, the data points corrected for the viscous drag are also shown by open circles. This correction does not alter the shape of the P-V curve obtained. Similar nearly straight P-V curves were obtained from 6 other beads with high  $P_o$  values (8.6-



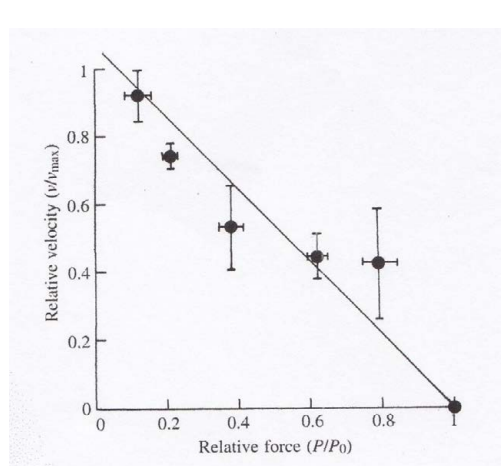
**Figure 10:** Constant velocity sliding of a bead, with cytoplasmic myosin attached, along actin cables under different positive loads, expressed relative to the maximum isometric force  $P_0$  (13 pN) [23].



**Figure 12:** Steady-state P-V relation of cytoplasmic myosin versus actin cable, constructed from 6 different beads, with large  $P_0$  values (8.6-13 pN). Bars indicate SD (n=6). Note that the P-V relation is straight [23].



**Figure 11:** Steady-state P-V curve of a bead, with cytoplasmic myosin attached, on actin cables (filled circles). The loads were applied in a random order. The amount of force (=load) is expressed relative to  $P_0$  (13 pN). Data points corrected for the viscous drag force (open circles) are also shown [23].



**Figure 13:** Steady-state P-V relation of cytoplasmic myosin versus actin cable, Constructed from 6 different beads with small  $P_0$  values (1.0-2.4 pN). Bars indicate SD (n=6) [23].

13 pN). Figure 12 shows averaged P-V relation constructed from 6 different beads with high  $P_0$  values [23].

Figure 13 is averaged P-V curve constructed from 6 different beads with small  $P_0$  values (1.0-2.4 pN), which may reflect extremely small numbers of cytoplasmic myosin molecules producing bead movement. Although the data points showed a large scatter, especially at high load region, the overall P-V curve was nearly straight [23].

### Kinetic Properties of Cytoplasmic Myosin Heads Producing the Ultra-fast Cytoplasmic Streaming

The hyperbolic P-V curve of contracting whole skeletal muscle is fitted to part of rectangular hyperbola as:

$$(P + a)V = b(P_0 - P),$$

where  $P$  is the load on muscle,  $P_0$  is the maximum isometric force generated by muscle, and  $a$  and  $b$  are constants. The P-V curve is generally characterized by values of the maximum unloaded shortening

velocity  $V_{max}$  and  $a/P_0$  [24]; the former determines its intercept with the velocity axis, while the latter determines its curvature. A large value of  $a/P_0$  is associated with a less prominent curvature of P-V curve, and a low efficiency of converting chemical energy derived from ATP hydrolysis into mechanical work [25]. On this basis, the nearly straight P-V curve of cytoplasmic myosin versus actin cable sliding (Figures 12 and 13) suggests a very low efficiency of the chemo-mechanical energy conversion in cytoplasmic myosin molecules. The value of  $V_{max}$  for cytoplasmic myosin versus actin cable sliding, on the other hand, is many times larger than that in skeletal and cardiac myosin versus actin cable sliding, suggesting that the maximum cycling rate of actin-myosin interaction is many times larger in the former than in the latter.

To explain the increasing degree of deviation of the P-V curves of skeletal and cardiac myosin versus actin cable sliding from the hyperbola with decreasing value of  $P_0$ , we used the concept of  $p$  and  $P_{off}$ . In contrast, the P-V curve of cytoplasmic myosin versus actin cable sliding was straight irrespective of the value of  $P_0$  (Figures 11 and 12) [23]. This implies that, in cytoplasmic streaming, the probability

of Poff is extremely small despite very small Po values. In other words, the value of  $p$ , representing the fraction of time in which a myosin head firmly attaches to actin to produce actin-myosin sliding, i.e. duty ratio [26], may be very large and close to unity. We think that the large duty ratio is essential for producing continuous cytoplasmic streaming with the ultra-high velocity.

### Possible Mechanism of the Ultra-high Cytoplasmic Streaming

Based on the concept of duty ratio, there are three basic kinetic factors, i.e. (1) duty ratio of myosin head ( $p$ ), (2) ATPase rate per myosin head ( $A$ ), and (3) unitary sliding distance produced by a myosin head ( $d$ ). The value of  $p$  is written as:

$$p = (d \times A) / V_{\max}$$

In the ATP-dependent actin-myosin sliding in skeletal muscle,  $p$  is  $\sim 0.05$  [27,28],  $A$  is  $\sim 20$ /s/head [29], and  $V_{\max}$  is  $\sim 3 \mu\text{m/s}$  [13]. Putting these values into the above equation,  $d$  is obtained to be  $\sim 8$  nm, a value consistent with the contraction model, in which myosin head power stroke is caused by change in myosin head configuration [30].

In the ATP-dependent actin-cytoplasmic myosin sliding,  $p$  is assumed to be 0.25, while  $V_{\max}$  is  $50 \mu\text{m/s}$  [23]. The value of  $d$  is unknown, but it is not  $>20$  nm, if the size of myosin head is taken into consideration. From these values, above equation indicates that  $A$  should be  $\sim 425$ /s/head. The extremely high cyclic rate of actin-cytoplasmic myosin interaction, however, contradicts the result that actin-activated ATPase activity of cytoplasmic myosin in solution is  $20$ /s/head, being similar to that in skeletal muscle myosin in solution [31].

The puzzling situation can be solved at least qualitatively by assuming a cooperative mechanical interaction between adjacent myosin heads extending from cytoplasmic myosin molecule attached to the same bead or the same cytoplasmic organelle. Since the value of  $p$ , i.e. duty ratio in cytoplasmic myosin head is close to unity, each cytoplasmic myosin head may have many chances of being pushed by adjacent myosin heads to result in rapid detachment from actin to restart its power stroke producing cytoplasmic streaming. This may markedly increase the cycling rate of actin-myosin interaction coupled with ATP hydrolysis in cytoplasmic myosin heads extending from the same bead or the same organelle.

This idea comes from the report that, if single skeletal muscle fibers are previously stretched to sarcomere length of  $3 \mu\text{m}$  with resting tension amounting  $\leq 7\%$  of  $P_o$ , the maximum shortening velocity of tetanized fibers, as measured by the slack test, increases  $\sim$ two fold [32]. As the resting tension decreases quickly at the beginning of applied release, the myosin heads attached to actin in tetanized fibers can be pushed in the direction of fiber shortening by the resting tension for an extremely short period of time. This possibility has already been discussed by us in a short review of cytoplasmic streaming [33].

### Conclusion

The large duty ratio of cytoplasmic myosin head might produce mechanical synchronization and summation of power strokes between adjacent myosin heads to result in marked increase in the rate of cyclic interaction to greatly increase the velocity of actin-myosin sliding. Considering the extremely small Reynolds number for myosin head power stroke in the cytoplasm, together with a duty ratio close to unity, adjacent myosin heads on the same cytoplasmic organelle can

effectively interact each other to produce the ultra-fast cytoplasmic streaming. Myosins are widely distributed in both animal and plant kingdoms, to comprise a superfamily, in which skeletal and cardiac myosins and cytoplasmic myosin are classified into myosin II and myosin XI, respectively [34]. To reach full understanding of myosin molecules, it is essential to study similarities and differences among different myosin types.

### Acknowledgement

We dedicate this article to the late Professor Eiji Kamitsubo for his contribution to our studies on the sliding between cytoplasmic myosin and actin cables using the centrifuge microscope.

### References

1. Kamiya N, Kuroda K (1956) Velocity distribution of the protoplasmic streaming in *Nitella* cells. *Bot Mag* 69: 544-554.
2. Kamitsubo E (1966) Motile protoplasmic fibrils in cells of Characeae. *Proc Japan Acad* 42: 640-643.
3. Nagai R, Rebhum I (1979) *J Ultrastruct Res* 14: 571-589.
4. Kersey YM, Wessels NK (1976) Localization of actin filaments in internodal cells of characean algae. A scanning and transmission electron microscope study. *J Cell Biol* 68: 264-275.
5. Kato T, Tonomura Y (1977) Identification of myosin in *Nitella flexilis*. *J Biochem* 82: 777-782.
6. Nagai R, Hayama T (1979) Ultrastructure of the endoplasmic factor responsible for cytoplasmic streaming in *Chara* internodal cells. *J Cell Sci* 36: 121-136.
7. Tazawa M, Shimmen T (1987) Cell motility and ionic relations in characean cells as revealed by internal perfusion and cell models. *Int Rev Cytol* 109: 259-312.
8. Williamson RE (1975) Cytoplasmic streaming in *Chara*: a cell model activated by ATP and inhibited by cytochalasin B. *J Cell Sci* 17: 655-668.
9. Williamson RE, Ashley CC (1982) Free  $\text{Ca}^{2+}$  and cytoplasmic streaming in the alga *Chara*. *Nature* 296: 647-651.
10. Tominaga Y, Shimmen T, Tazawa M (1983) Control of cytoplasmic streaming by extracellular  $\text{Ca}^{2+}$  in permeabilized *Nitella* cells. *Protoplasma* 116: 75-77.
11. Shimmen T, Yano M (1984) Active sliding movement of latex beads coated with skeletal muscle myosin on *Chara* actin bundles. *Protoplasma* 121: 132-137.
12. Sheetz MP, Spudich JA (1983) *Nature* 303: 31-35.
13. Oiwa K, Chaen S, Kamitsubo E, Shimmen T, Sugi H (1990) Steady-state force-velocity relation in the ATP-dependent sliding movement of myosin-coated beads on actin cables in vitro studied with a centrifuge microscope. *Proc Natl Acad Sci USA* 87: 7893-7897.
14. Higashi-Fujime S, Ishikawa R, Iwasawa H, Kagami O, Kurimoto E, et al. (1995) The fastest-actin-based motor protein from the green algae, *Chara*, and its distinct mode of interaction with actin. *FEBS Lett* 375: 151-154.
15. Yamamoto Y, Hamada S, Kashiwayama T (1999) *Cell Mol Life Sci* 56: 226-232.
16. Tsuchiya T, Tanaka H, Shirakawa I, Karr T, Sugi H (1998) Evidence for the essential role of myosin subfragment-2 in the ATP-dependent actin-myosin sliding in muscle contraction. *Jpn J Physiol* 48: 383-387.
17. Edman KAP (1988) Contractile performance of skeletal muscle fibres. *J Physiol* 484: 301-321.
18. Iwamoto H, Sugaya R, Sugi H (1990) Force-velocity relation of frog skeletal muscle fibres shortening under continuously changing load. *J Physiol* 422: 185-202.
19. Sugi H, Kobayashi T, Gross T, Noguchi K, Karr T, et al. (1992) Contraction characteristics and ATPase activity of skeletal muscle fibers in the presence of antibody to myosin subfragment 2. *Proc. Natl. Acad. Sci. USA* 89: 6134-6137.
20. Ohno T, Arai T, Chaen S, Sugi H (2017) *J Nanomed Nanotechnol* 8: 1000428.
21. Sugiura S, Yamashita H, Sata M, Momomura S, Serizawa T, et al. (1995) Force-velocity relations of rat cardiac myosin isozymes sliding on algal cell actin cables in vitro. *Biochim. Biophys Acta* 1231: 69-75.

22. Huxley AF (1957) Muscle structure and theories of contraction. *Prog Biophys Biophys Chem* 7: 255-318.
23. Chaen S, Inoue J, Sugi H (1995) The force-velocity relationship of the ATP-dependent actin-myosin sliding causing cytoplasmic streaming in algal cells, studied using a centrifuge microscope. *J Exp Biol* 198: 1021-1027.
24. Woledge RC, Curtin NA, Homsher E (1985) *Energetic Aspects of Muscle Contraction*, Academic Press, London
25. Woledge RC (1968) The energetics of tortoise muscle. *J Physiol* 197: 685-707.
26. Howard J (1997) Molecular motors: structural adaptations to cellular functions. *Nature* 389: 561-567.
27. Ueyda TPQ, Kron SJ, Spudich JA (1990) Myosin step size: estimation from slow sliding movement of actin over low densities of heavy meromyosin. *J Mol Biol* 214: 699-710.
28. Finer JT, Simmons RM, Spudich JA (1994) Single myosin molecule mechanics: piconewton forces and nanometre steps. *Nature* 368: 113-119.
29. Toyoshima Y, Kron SJ, Spudich JA (1990) The myosin step size: measurement of the unit displacement per ATP hydrolyzed in an in vitro assay. *Proc Natl Acad Sci USA* 87: 7130-7134.
30. Huxley AF, Simmons RM (1971) Proposed mechanism of force generation in striated muscle. *Nature* 233: 533-538.
31. Tanimura A, Higashi-Fujime S (2002) Force-velocity relationships in actin-myosin interactions causing cytoplasmic streaming in algal cells. *J Muscle Res Cell Motil* 23: 185.
32. Edman KAP (1979) The velocity of unloaded shortening and its relation to sarcomere length and isometric force in vertebrate muscle fibres. *J Physiol* 291: 143-159.
33. Sugi H, Chaen S (2003) Force-velocity relationships in actin-myosin interactions causing cytoplasmic streaming in algal cells. *J Exp Biol* 206: 1971-1976.
34. Sellers J (2000) Myosins: a diverse superfamily. *Biochim Biophys Acta* 1496: 3-22.